

REVIEW

Open Access



# Reprogramming of sentinel lymph node microenvironment during tumor metastasis

Yen-Liang Li<sup>1</sup> and Wen-Chun Hung<sup>1,2\*</sup>

## Abstract

Metastasis is a major cause of death in patients with cancer. The two main routes for cancer cell dissemination are the blood and lymphatic systems. The underlying mechanism of hematogenous metastasis has been well characterized in the past few decades. However, our understanding of the molecular basis of lymphatic metastasis remains at a premature stage. Conceptually, cancer cells invade into lymphatic capillary, passively move to collecting lymphatic vessels, migrate into sentinel lymph node (SLN; the first lymph node to which cancer cells spread from the primary tumor), and enter the blood circulatory system via the subclavian vein. Before arriving, cancer cells release specific soluble factors to modulate the microenvironment in SLN to establish a beachhead for successful colonization. After colonization, cancer cells inhibit anti-tumor immunity by inducing the recruitment of regulatory T cell and myeloid-derived suppressor cells, suppressing the function of dendritic cell and CD8<sup>+</sup> T cell, and promoting the release of immunosuppressive cytokines. The development of novel strategies to reverse cancer cell-triggered SLN remodeling may re-activate immunity to reduce beachhead buildup and distant metastasis. In addition to being a microanatomic location for metastasis, the SLN is also an important site for immune modulation. Nanotechnology-based approaches to deliver lymph node-tropic antibodies or drug-conjugated nanoparticles to kill cancer cells on site are a new direction for cancer treatment. Conversely, the induction of stronger immunity by promoting antigen presentation in lymph nodes provides an alternate way to enhance the efficacy of immune checkpoint therapy and cancer vaccine. In this review article, we summarize recent findings on the reprogramming of SLN during lymphatic invasion and discuss the possibility of inhibiting tumor metastasis and eliciting anti-tumor immunity by targeting SLN.

**Keywords:** Lymph node, Immune, Metastasis, Microenvironment

## Introduction

Cancer cells are highly proliferating cells that develop in heterogeneous environments. When tumors grow to a certain size, cancer cells face with oxygen/nutrient supply and metabolic waste accumulation. Escape from primary tumors (known as metastasis) is a selection pressure or an advantage for cancer cells to counteract the harmful microenvironment. Metastasis is a biological process that guarantees the sustained growth of cancer cells. It

remains largely incurable and is responsible for up to 90% of cancer-associated mortality. The invasion-metastasis cascade of cancer cells is orchestrated by the following events: (1) invasion into the surrounding tissues containing abundant stromal cells and dense extracellular matrix (ECM); (2) intravasation into the circulation systems including the blood or lymphatic system; (3) survival in the circulation and travel to distant organs; (4) extravasation from blood vessels for colonization; (5) establishment of micrometastases to build a beachhead at the colonized sites; and (6) expansion in the new microenvironment to generate a secondary tumor.

\*Correspondence: [hung1228@nhri.edu.tw](mailto:hung1228@nhri.edu.tw)

<sup>1</sup> National Institute of Cancer Research, National Health Research Institutes, Tainan 704, Taiwan  
Full list of author information is available at the end of the article



© The Author(s) 2022. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

### **Metastatic spread via blood and lymphatic vessels**

Cancer cells are mainly disseminated via the blood and lymphatic systems. The lymphatic system not only serves as a circulation system for the collection of interstitial fluid into the bloodstream [1], but also as an immune defense barrier to ensure the cleanliness of lymph fluid returning to the circulation. Terminal lymphatic vessels are thin-walled capillaries without pericyte coverage that are easier for cancer cells to penetrate. It has been proposed that approximately 95% of peritumoral vessels invaded by cancer cells are lymphatics [2, 3]. The lymphatic vasculature comprises lymphatic endothelial cells (LECs) that express molecular markers including lymphatic vessel endothelial receptor 1 (LYVE-1), prospero homeobox protein 1 (PROX1), podoplanin (PDPN), vascular endothelial growth factor receptor-3 (VEGFR-3), neuropilin-2 (NRP-2), and C-C motif chemokine ligand 21 (CCL21). Vascular endothelial growth factor-A (VEGF-A), -C, and -D released by cancer cells stimulate the growth of peripheral tumor lymphatic vessels and promote the invasion of cancer cells into nearby lymph nodes (known as sentinel lymph node, SLN) to enhance tumor metastasis [4–7]. Accumulating evidence suggests that VEGFR-3-mediated activation of LECs is a crucial step in the induction of lymphatic metastasis [8]. However, other factors, such as lymph flow rate, surface receptors expressed on tumor cells, and chemokines released from LECs also affect the entry of cancer cells to lymphatic vessels [9–11]. It is possible that these factors work together to promote lymphatic metastasis.

### **The importance of lymph node invasion in tumor metastasis—a question still under debate**

Animal studies have strongly supported the role of lymphangiogenesis and lymph node invasion in tumor metastasis. The orthotopic transplantation of VEGF-C-overexpressing breast cancer cells onto nude mice increased intratumoral lymphangiogenesis and significantly promoted tumor metastasis to the regional lymph nodes and lungs [12, 13]. Similarly, VEGF-A has been observed to trigger SLN lymphangiogenesis and lymphatic metastasis to enhance tumor spread [14, 15]. The ectopic expression of other lymphangiogenic factors, such as VEGF-D, platelet-derived growth factor-BB, and fibroblast growth factor-2, also induced metastasis in different cancer models [16–18]. In clinical setting, micrometastasis in SLN is a crucial factor associated with reduced distant metastasis-free survival and overall survival in cutaneous melanoma [19]. Leiter et al. also showed that dissection of SLN in primary melanoma decreased distant metastasis [20]. Scoring of immune and stromal features of SLN predicted distant metastasis in breast cancer patients [21]. In addition, lymph node

metastasis increased the incidence of distant metastasis (hazard ratio=3.495) in thyroid cancer [22]. By studying somatic variants in specific DNA regions to address the origins of lymphatic and distant metastasis, Naxerova et al. demonstrated that 36% of distant metastasis arose from lymph nodes in colorectal cancer [23]. Similarly, phylogenetic investigation also showed that 25% of metastatic tumors at distant sites were derived from lymph node metastasis [24]. A very recent study clearly demonstrated that colonization of lymph nodes by cancer cells elicited a chronic interferon signaling and triggered antigen-specific immune tolerance to promote distant metastasis [25]. However, several clinical intervention studies showed that prophylactic LN removal does not improve overall survival in melanoma [26], thyroid cancer [27], and breast cancer [28]. A long-term follow-up of a randomized trial demonstrated that localized lymph node metastasis, distant metastasis and patient's survival were not strongly correlated [29]. In addition, lymph node-negative colorectal cancer patients had higher incidence of lung metastasis [30]. In contrast, a prospective multicenter study of early stage endometrial cancer suggested that SLN biopsy provides important information to for tailoring adjuvant therapy [31]. SLN biopsy continues to be a critical procedure in the clinical management of patients with salivary gland tumors [32]. It should be noted that clinical outcome of lymph node-positive patients is often worse than that of lymph node-negative patients in some types of human cancer. For instance, in clinical stage IIB and IIC melanoma patients, SLN status is the most important prognostic factor and positive SLN involvement is strongly correlated reduced distant recurrence-free survival and disease-specific survival [33]. Similar finding was reported in a retrospective cohort of 2086 melanoma patients [34]. In addition, a national cohort study in which 8562 patients were included also concluded that SLN status is a critical prognostic factor in stage IIB/C melanoma patients [35]. In colon cancer, the average survival of lymph node-positive patients was shorter than that of lymph node-negative patients (66 vs. 89 months) [36]. In addition, the number of positive lymph nodes affected the response of patient to chemotherapy. The average survivals of patients with 1-, 2-, and 3-positive lymph nodes who received chemotherapy were 108, 83 and 54 months respectively. In pancreatic cancer, lymph node-negative patients have a longer median overall survival than lymph node-positive patients (25 vs. 16 months) [37]. In esophageal cancer, lymph node-positive patients with tumors localized at middle and lower regions had a worse prognosis than the lymph node-negative patient [38]. Emerging evidence suggests that tumor metastasis may occur at a very early stage of tumorigenesis. Therefore, resection of the primary tumor and SLN

may not significantly improve patient survival if cancer cells have already been seeded on distant organs via the hematological and lymphatic systems. As aforementioned in the studies of evolutionary relation between primary tumor, lymph node metastasis and distant metastasis [23, 24], around 30–40% of distant metastasis arose from lymph node metastasis. Many cancers may metastasize to distant organs via blood and lymphatic systems simultaneously and the contribution of lymph node metastasis to distant dissemination could be cancer type-dependent. Moreover, other important factors including the number of lymph nodes evaluated, the number of positive lymph nodes, the features of lymph node microenvironment and the depth of lymph node involvement all affect the results of pathological evaluation and the conclusions of clinical association. For example, a systematic review of a total 61,371 colon cancer patients showed that the number of surgically dissected lymph nodes evaluated was positively correlated with the survival of stage II and III patients [39]. Enhancement of angiogenesis and lymphangiogenesis in the SLN was found to be linked with distant metastasis and survival of melanoma patients [40]. In addition, the number of B cells in the SLN, regardless of the status of cancer cell invasion, also predicted disease-free survival in patients with breast cancer [41]. Therefore, the importance of lymph node invasion in the induction of distant metastasis warrants continuous study.

### Lymph node structure

The lymphatic system comprises a large network of lymph and lymphatic capillaries, collecting lymphatic vessels, lymph nodes and lymphoid organs. Lymph, the fluid that drains from cells and tissues, contains small molecules (minerals and amino acids), large molecules (proteins and lipids), and cells (damaged cells or immune cells). It flows from the lymphatic capillary, the terminal vessel of the lymphatic network, to a large collecting lymphatic vessel that further connects to the lymph node, a kidney-shaped organ of the lymphatic system. Lymph nodes are classified as "secondary" lymphoid organ, while the primary lymphoid organs comprise the thymus, tonsils, spleen, and bone marrow. Approximately thousands

of lymph node are linked throughout the body by lymphatic vessels [42] and are particularly distributed in the chest, neck, pelvis, axilla, and inguinal region, and in association with the blood vessels of the intestines [43]. The anatomical structure of the lymph node is divided into several compartments. The outer portion of the lymph node consists of the cortex, containing the B-cell follicle, and the paracortex, containing the T-cell zone. The inner portion of the node is the medulla which contains blood vessels, sinuses, and medullary cord. Antibody-producing plasma cells, macrophages, and B cells are the major cell types in the medullary cord. A specialized structure, high endothelial venules (HEVs), found in the paracortex are the main routes for lymphocytes to enter the lymph node.

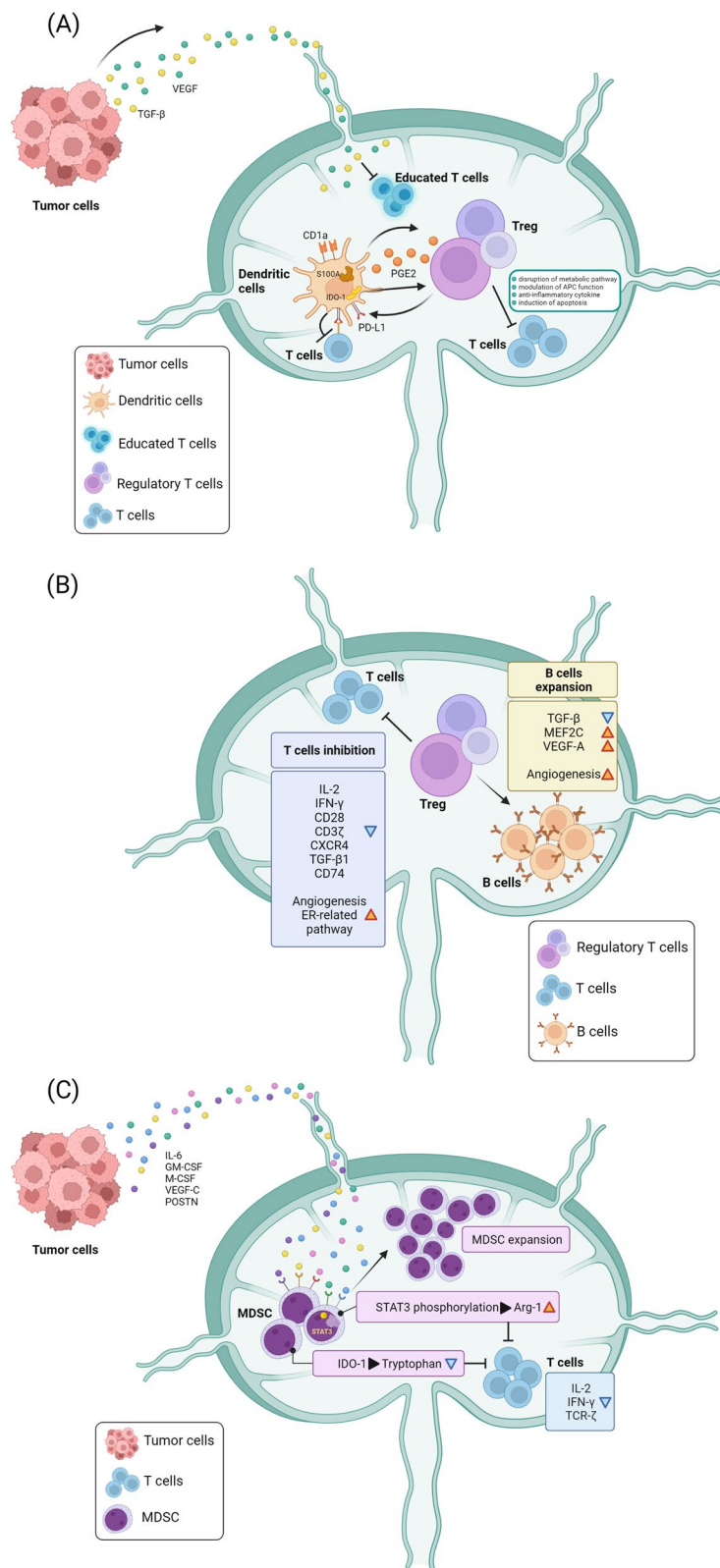
The SLN is defined as the first lymph node with direct lymphatic flow from the primary tumor and is the beachhead for the earliest stage of lymphatic metastasis. Clinically, the presence of tumor cells in the SLN is a prognostic factor associated with cancer progression and poor patient outcome [44–46]. The biological features of SLN include the enhancement of lymphangiogenesis, increase in lymph flow [47], structural remodeling of HEVs [4, 48], enhanced recruitment of myeloid cells, and reduction of effector lymphocytes [49], all of which contribute to the establishment of a pre-metastatic microenvironment for the entry and survival of cancer cells.

### Reprogramming in immune cells in the SLN

The immune microenvironment in the lymph node is orchestrated by immune cells including macrophages, dendritic cells (DCs), T cells, B cells, and non-immune cells, such as fibroblastic reticular cells (FRCs), blood endothelial cells (BECs), and LECs. One of the key regulators of the host immune system to attack cancer cells are DCs, highly specialized antigen-presenting cells, that play a crucial role in the initiation of cellular immunity (Fig. 1A). Previous studies have demonstrated that the anti-cancer activity of T cells is dramatically attenuated in the absence of DC [50–53]. In the SLN, DC-induced T-cell activation is significantly impaired by cancer cells via direct cell–cell contact or cancer cell-secreted

(See figure on next page.)

**Fig. 1** Immune microenvironment modulation in the SLN during cancer progression. **A** Tumor cell-derived soluble factors including TGF- $\beta$  and VEGF impair T cell and lymphocyte activation by DCs. Expression of COX2 and IDO-1 in DCs also contribute to the expansion of Treg cells which directly diminish T cell activity. Treg cells stimulate a feedback loop to enhance PD-L1 expression on DCs, leading to immune suppression of T cells via the receptor PD-1. S100 and CD1a elevated in the DCs in the SLN also modulate antigen presentation of DCs. **B** After expansion, Treg cells enter the SLN via HEV and lymphatic vessels. Treg cells inhibit T cell activity, decrease pivotal molecules such as IL-2, IFN- $\gamma$ , CD28, CD3- $\zeta$  chain, CXCR4, TGF- $\beta$ 1 and CD74 to suppress the proliferation, activation and differentiation of T cells. Treg cells also induce B cells expansion and enhance MEF2C expression to alter the proliferation and survival of B cells. In addition, Treg cells promote the estrogen receptor-related pathways in T cells to facilitate angiogenesis and lymphangiogenesis. **C** MDSCs are recruited to the SLN and are activated by external signals such as IL-6, GM-CSF, M-CSF, VEGF-C and POSTN released from distant tumor cells. STAT3 phosphorylation in MDSCs stimulate the production of Arg-1 to compete the essential amino acid with T cells and to suppress T cell function. In addition, IDO-1-expressing MDSCs catalyze the metabolism of tryptophan to kynurenine, a metabolite which induces T cell apoptosis



**Fig. 1** (See legend on previous page.)



factors, such as transforming growth factor- $\beta$  (TGF- $\beta$ ) and VEGF. Munn and Sharma et al. showed that a small population of indoleamine 2,3-dioxygenase 1 (IDO1)-expressing plasmacytoid DCs in the SLN was capable of inducing regulatory T-cell (Treg) generation and T-cell anergy, which was linked with decreased T-cell response to tumor antigens [54, 55]. Sakakura et al. also demonstrated that the increase in S100<sup>+</sup> and CD1a<sup>+</sup> DCs in the SLN of patients with oral cancer suppresses immune response [56]. In addition, DCs have been reported to produce cyclooxygenase-2-derived prostaglandin E2 to promote the accumulation of Tregs in the SLN [46].

Lymphocytes are recruited into the lymph node mainly via the HEVs and lymphatic vessels. A previous study demonstrated a marked decrease in the number of CD4<sup>+</sup> T helper (Th) cells in the advanced clinical stage of melanoma [57]. A reduction in lymphocyte infiltration in head and neck cancers has also been reported [56]. In addition, lymphocytes in the SLN of oral cancer were found to express distinct immune molecules, suggesting phenotypic alterations in these cells [58]. In parallel, another important sub-population is the immunosuppressive CD4<sup>+</sup>CD25<sup>hi</sup> forkhead box P3 (FOXP3<sup>+</sup>) Tregs (Fig. 1B). These predominant cells are significantly elevated in tumor cell-positive lymph node in patients with different cancers [59, 60]. Tregs inhibit the proliferation of CD8<sup>+</sup> T cells and weaken the ability of lymphocytes to produce interleukin (IL)-2 and interferon (IFN)- $\gamma$ . The heterogeneity of T cells in primary tumor and SLN is very significant. Only a very small number of expanded T cells have been found in the SLN [48, 49]. Additionally, CD4<sup>+</sup> T cells were dramatically decreased in the SLN in breast cancer [61, 62]. A similar finding has also been reported in oral cancer [58]. The accumulation of B cells in the SLN has been found to be associated with lymphangiogenesis and increased lymph flow, thereby effectively promoting the dissemination of lymphomas and solid tumors [63]. The increase in B cells in patients with cancer indicated the activation of clonal expansion, probably triggered by the recognition of tumor antigens, and suggested the enhancement of the apoptosis-inducing ability of B cells [64, 65].

Recently, we performed single-cell RNA sequencing to explore alterations in gene expressions in different cell populations in the SLN and identified the molecular pathways altered in T cells in the SLN [66]. Our results showed that angiogenesis-related gene sets were significantly upregulated in Cd4<sup>+</sup>, and Cd8<sup>+</sup> T-cells and Tregs. Consistent with our findings, a recent study demonstrated an increase in the proportion of T cells, B cells, DCs and natural killer (NK) cells in metastatic lymph nodes [67]. Interestingly, the accumulation of N2 type neutrophil (Cd54<sup>low</sup>) was significantly enhanced.

Moreover, the estrogen response gene signature, which is involved in enhancing breast cancer progression, was elevated in Cd4<sup>+</sup> T cells in the SLN. Estrogen receptor 1 (Esr1), a transcription factor that binds directly to the gene promoter of retinoic acid-receptor-related orphan nuclear receptor  $\gamma$ , may suppress the differentiation and function of Th17 cells [68]. Moreover, Esr1 may inhibit follicular helper T-cell activation to prevent autoimmunity [69]. Our finding that Esr1 signaling is activated in Cd4<sup>+</sup> T cells suggests a suppressed immunity in the SLN. Clusters of differentiated genes by gene set enrichment analysis using molecular signatures database revealed that the gene sets related to Foxp3-mediated Treg transcriptional regulation in the C7 immunological signature were altered, suggesting the enhancement of Treg activation and generation of an immune suppressive environment in the SLN. The expression of genes related to proliferation (Cxcr4), polarization (Cxcr4, Tgfb1, and Cd4), and differentiation (Cd74 and Cd4) in Cd4<sup>+</sup> T cells in the SLN was reduced. The differentiation marker Cd74 was also downregulated in Cd8<sup>+</sup> T cells in the SLN. Our results also demonstrated enhancement of the angiogenesis pathway in B cells, consistent with previous findings that tumor-associated B cells contribute to tumor progression by stimulating angiogenesis [70]. An increase in myocyte-specific enhancer factor 2C (Mef2c) in B cells was found in the SLN. Mef2c is a key transcription factor that increases B-cell proliferation and survival [71, 72]. In addition, this molecule protects B cell lymphopoiesis under stress conditions by regulating B-cell specific gene expression [73]. Finally, TGF- $\beta$ , a critical cytokine in controlling the development of B cells from pre-B cells to immunoglobulin-secreting plasma cells [74], was downregulated in the SLN, indicating inhibition of functional differentiation.

Myeloid-derived suppressor cells (MDSCs) also participate in the enhancement of tumor growth and metastasis by accumulating in the SLN to suppress immune reactivity (Fig. 1C). Cancer cells secrete IL-6, VEGF, macrophage colony-stimulating factor (M-CSF), and granulocyte-M-CSF (GM-CSF) to promote MDSC expansion and enhance their recruitment to SLN [75, 76]. MDSCs influence immune responses by (1) inducing the development and expansion of Tregs [77–79]; (2) depriving amino acids that are essential for T-cell growth and differentiation [80, 81]; (3) releasing oxidizing molecules, including hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) and peroxynitrite (ONOO<sup>-</sup>) to increase immune cell apoptosis [82, 83], and (4) interfering with T-cell migration [84, 85]. By increasing IDO1 expression, MDSCs decrease the immune response of T cells and trigger T-cell apoptosis via kynurenine production [86, 87]. These studies highlight the role of MDSCs in immune suppression and suggest the possibility of

targeting MDSCs to overcome the immune escape. Macrophages are another myeloid lineage cell considered to be involved in lymph node metastasis. Broadly speaking, there are two main groups of macrophage designated as M1 and M2. M1 macrophages release inflammatory cytokines, whereas M2 macrophages exhibit anti-inflammatory activity. Tumor-associated macrophages (TAM) acquire an M2 phenotype that contributes to tumor growth and progression. It had been demonstrated that TAM level is significantly associated with pathologically positive-lymph node and is linked with enhanced lymphangiogenesis in the SLN [88]. In various types of cancer, the reduced expression of a unique type of CD169<sup>+</sup> macrophages in the SLN has been correlated with poor clinical outcome [89–91]. Collectively, existing data suggest that the immune microenvironment in the SLN is conditioned prior to cancer cell arrival, and re-activation of anti-cancer immunity in the SLN may prevent tumor metastasis.

#### Reprogramming in BECs/HEVs in the SLN

The vascular endothelium plays a central role in the regulation of oxygen/metabolite exchange and recruitment of immune cells to lymphoid tissues. The vascular system of lymph node in mammals consists of arteries, capillaries, post-capillary venules and vein [92]. Recently, two transcriptome analyses of different murine organs revealed molecular markers of BECs in lymph nodes according to their location [93, 94]. HEVs are specialized post-capillary vessels with high expression of peripheral node addressin (PNAd), Ccl21 and Cd105 [95, 96]. These molecules are important for the selective recruitment of lymphocytes to the lymph nodes. Loss of PNAd in metastatic lymph nodes has been shown to affect lymphocyte homing [4]. Lymphocyte recruitment into the SLN occurs mainly through HEVs and partially from the draining afferent lymphatic vessels. The remodeling of HEVs (Fig. 2) in the SLN decreases the recruitment of lymphocytes and promotes the establishment of a pre-metastatic niche [97]. Studies in human samples have found reduced vessel wall thickness and increased vessel diameter in HEVs in the SLN [4, 98, 99].

VEGF plays a pivotal role in stimulating BEC proliferation. This angiogenic factor not only acts as a survival factor for endothelial cells, but also stimulates them to degrade the ECM for sprouting and migration. Therefore, structural and functional alterations in the BECs of the SLN also accelerate lymphatic metastasis (Fig. 3).

#### Reprogramming in LECs in the SLN

LECs are defined as a specialized population of endothelial cells that comprise lymphatic vessel in lymph node, which are faithfully characterized by PDPN and LYVE-1

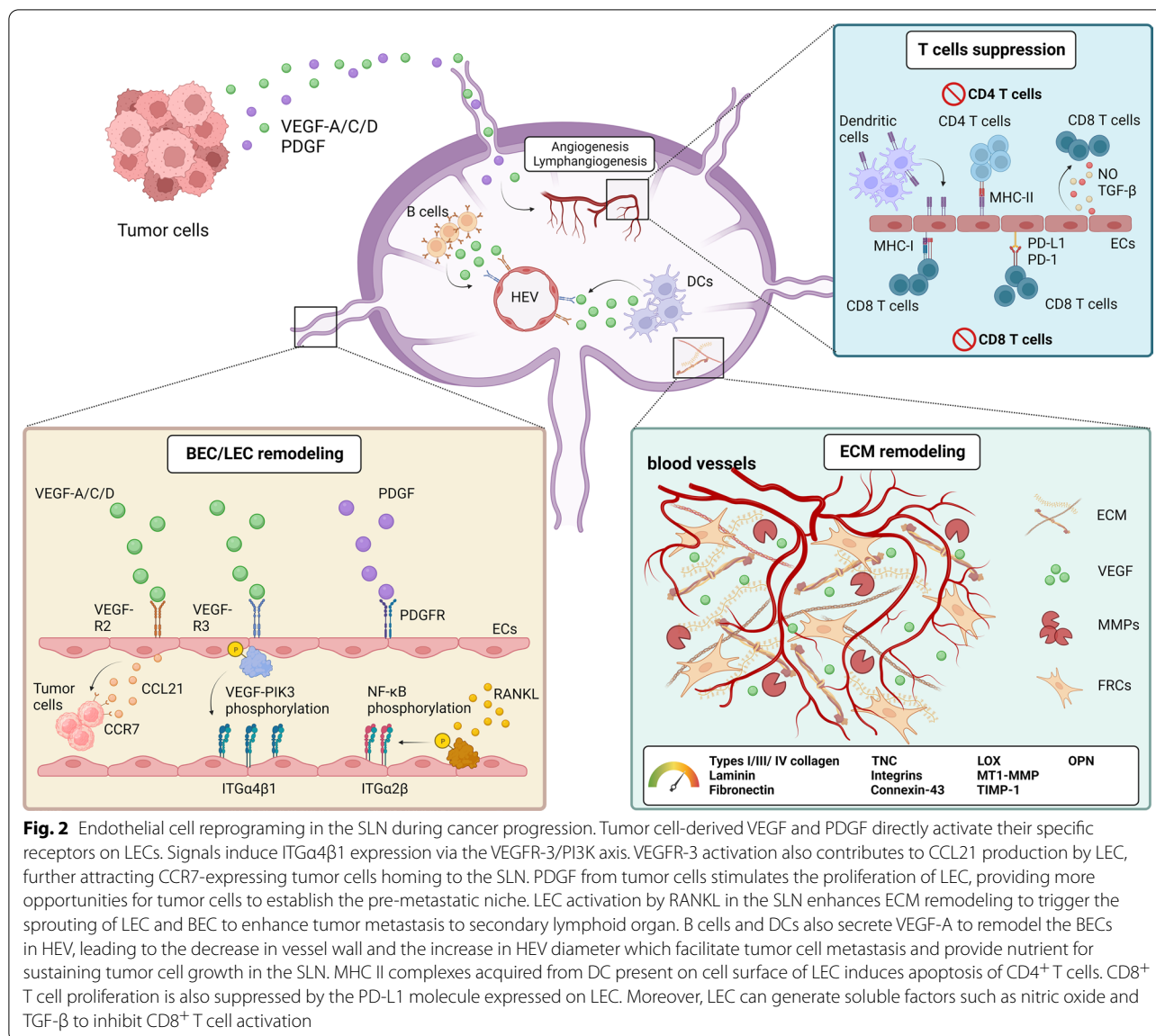
and are localized to the subcapsular, cortical, and medullary sinuses [100–102]. Among the pro-lymphangiogenic factors, VEGF-C exhibits the most potent activity in inducing lymphangiogenesis, and VEGFR3 on LECs is the major receptor involved in tumor-associated lymphangiogenesis and lymphatic metastasis [102, 103]. Recently, single-cell transcriptomic analysis revealed different subtypes of murine LECs in skin-draining lymph nodes, suggesting the complexity of their function [104].

Currently, VEGF-A and VEGF-C are the most well-documented soluble factors correlated with the establishment of a pre-metastatic microenvironment in the SLN [5, 105]. The remodeling of LECs via the VEGF-C-PI3K axis is critical for tumor-associated lymphangiogenesis. This signaling pathway enhances integrin  $\alpha\beta 1$  expression on LECs to attract Vcam-1-expressing tumor cells [106]. Simultaneously, VEGF-C also enhanced the expression of Ccl21 in the lymphatic endothelium to promote the entry of Ccr7<sup>+</sup> cancer cells into the SLN [107]. In contrast, lymphangiogenesis has been found to increase the lymph flow rate, which also accelerates lymph node metastasis [49, 108–110]. LECs in the SLN also expressed higher levels of receptor activator of nuclear factor kappa-B (Rank), and stromal reticular cells activated LECs via RANK ligand (Rankl) to induce LEC remodeling [111].

The functional reprogramming of LECs also affects their immunomodulatory activities. Under physiological circumstance, LECs may present a variety of peripheral tissue antigen on major histocompatibility complex (MHC) class I molecules to induce immune tolerance and modulate Cd8<sup>+</sup> T cell proliferation through programmed death protein 1 (Pd-1)/programmed death-ligand 1 (Pd-1) signaling [112]. In the SLN, by presenting soluble tumor-associated antigens from the lymph, LECs could trigger dysfunction in Cd8<sup>+</sup> T cell and increase T-cell apoptosis by regulating the expression of Pd-1, Cd80 and cytotoxic T-lymphocyte associated protein 4 (Ctla4) [112]. In addition, LECs express MHC class I on the surface, leading to functional alteration of Cd8<sup>+</sup> T cells [53, 113]. Moreover, LECs may acquire peptide/MHC II complexes from DC and present on their cell surface to trigger apoptosis of Cd4<sup>+</sup> T cells [112]. Moreover, LECs can present tumor antigens and produce immunosuppressive molecules such as kynurenine, nitric oxide, and TGF- $\beta$  to construct an immunosuppressive microenvironment in the SLN [53, 112–118].

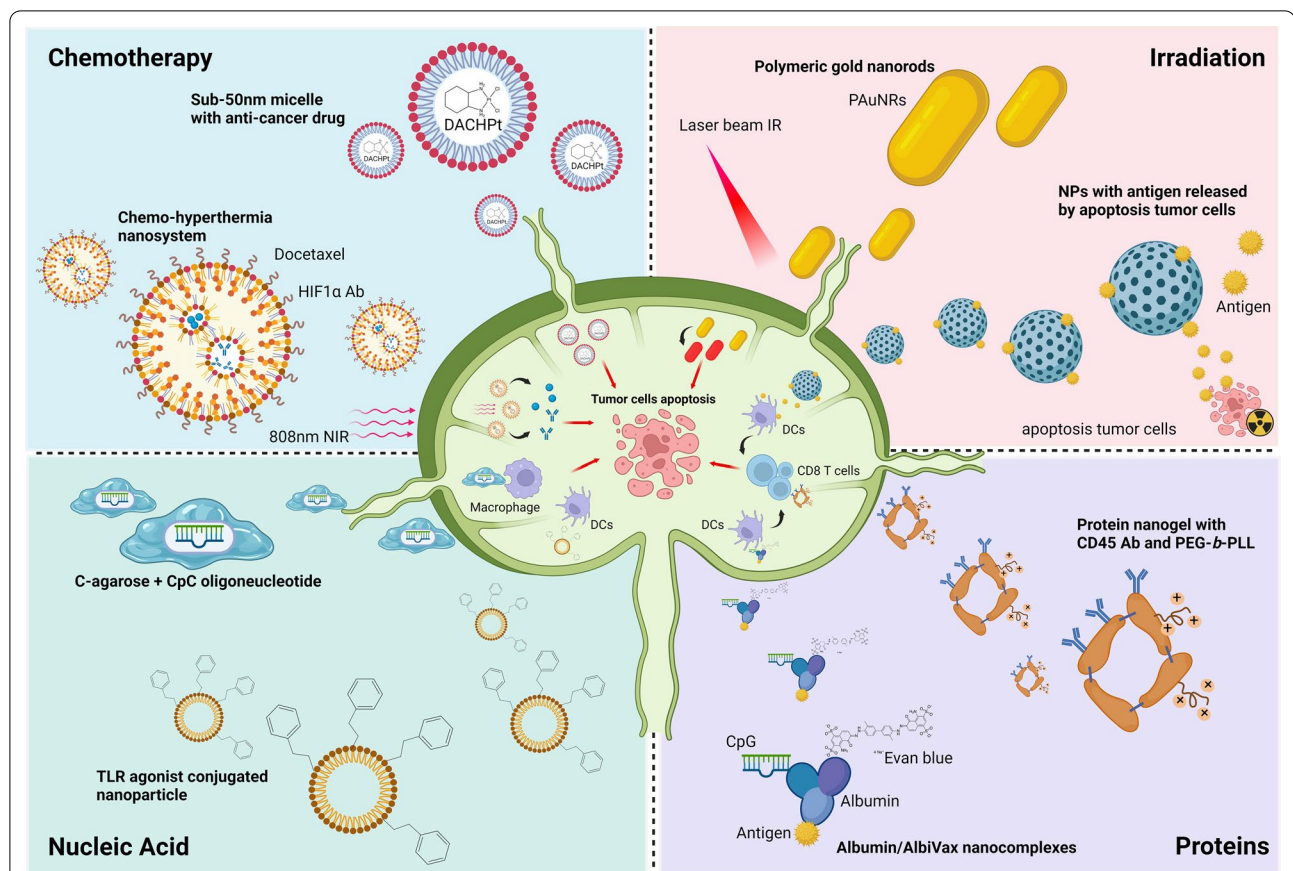
#### Reprogramming in stromal cells in the SLN

Lymph node stromal cells comprise distinct cell types: lymphoid tissue organizer (LTo), follicular DCs (FDCs), FRCs, marginal reticular cell (MRCs), integrin  $\alpha 7$  pericytes (IAPs), BECs, and LECs. LTos recruit



hematopoietic cells to lymph node. FDCs found in the cortex around the B cell zone of lymph node are important for supporting B cell survival. FRC produce various ECM proteins including fibrillary type I and III collagen, collagen type IV, laminin, fibronectin, tenascin-C, and integrins to form a reticular venue for immune cell movement and to strengthen the lymph node structure [119, 120]. FRCs play a crucial role in maintaining HEV integrity and immune cell survival. MRC constitutively produces chemokine CXCL13 to modulate the characteristic and functionality of LTo cells. IAP is a newly identified type of stromal cells located around blood vessels in the lymph node with uncharacterized biological function.

The reprogramming of stromal cells in the SLN has been previously reported in a B16.F10 melanoma mouse model [121]. This study demonstrated that genes participating in diverse biological processes, including growth, metabolism, mitochondrial function, cell motility, and cell–cell junction, were upregulated in FRC in the SLN of tumor-bearing mice when compared to naïve lymph node. Both gene set enrichment analysis and interpretative phenomenological analysis identified upregulated expression of genes encoding chemokines, cytokines and their downstream mediators in the SLN while several factors such as IL-19, IL-7, CCL4, and CCL21 were downregulated in comparison with those in naïve lymph node. Transcriptional profiling



**Fig. 3** Novel LN-targeted therapeutic strategies. Nanoparticles have been developed to kill tumor cells or re-activate antitumor activity in the LN. For chemotherapy, 808 nm NIR-triggered nanosystem achieves synergistic chemo-hyperthermia effects to eliminate tumor cells in the metastatic lymph nodes. Nanocarriers can bring nucleic acid toward the LN. CpG oligodeoxynucleotides equipped with C-agarose display high affinity to the macrophages in the lymph node sinus and effectively trigger anti-tumor immune responses in the LN. TLR agonist-conjugated nanoparticles activate DCs in the LN and stimulate T cell activity. In radiotherapy, gold nanorods under short-term NIR laser irradiation may increase tumor cell apoptosis via a thermodynamic effect. Tumor antigens released from cancer cells after IR treatment can be utilized as cargos and carried by nanoparticles to the LN, thus enhancing antigen presentation by DCs to activate T cells. Nanogels carrying an IL-15 superagonist complex coated with CD45 antibody can bind to CD8<sup>+</sup> T cells specifically and efficiently stimulate the proliferation of T cells by IL-15 stimulation. On the other hand, by conjugating albumin-binding vaccines with Evans blue, this nanocomplex provides another way for vaccine delivery and cancer immunotherapy

also showed that the expression of activation markers, including PDPN, fibronectin 1, Cd248, actin  $\alpha$ 2, S100A4, vimentin, myosin light chain, and collagens, was enhanced, indicating the activation of FRCs in the SLN.

Based on our single-cell RNA sequencing data, FRC in the SLN expressed a strong elevation of the oxidative phosphorylation (OXPHOS) pathway signature [55]. Twenty-seven genes of complex I, one of complex II, six of complex III, nine of complex IV, and four of complex V of the respiratory chain were differentially expressed, suggesting massive ATP consumption in the SLN. These data suggest that a metabolic switch in the SLN promotes lymphatic metastasis.

### The mediators for SLN reprogramming

#### Growth factors and chemokines

Mounting evidence suggests that distinct chemokine-receptor signaling pathways contribute to the trafficking of cancer cells to the lymph node (Table 1). Compared to naïve lymph nodes, the expression of GM-CSF, IFN- $\gamma$ , IL-2, and IL-10 is elevated in the SLN in melanoma [122, 123]. In breast cancer, the expression of CD83, IL-12p40, IFN- $\gamma$ , IL-10, and Foxp3 was evidently upregulated in the tumor-infiltrated SLN [124]. In non-small-cell lung cancer, tumor-derived TGF- $\beta$  reduces the number of DCs in the SLN [53]. Hirakawa et al. reported that primary tumors overexpressed VEGF-A to induce lymphangiogenesis in the



**Table 1** Cytokines and chemokines modulation in SLN during cancer progression

Cancer type	Proteins and nucleic acids	Effect	Refs.
Cytokines	COX-2	Macrophages phenotype changes	
	GM-CSF	Proliferation and differentiation of DCS	[5]
Melanoma	IFN- $\gamma$	Production of cytotoxic cells	[122]
	IL-10	Dampens acquired Th1 and Th2 cell cytokine production	[123]
	VEGF-A	Lymphangiogenesis	
Breast cancer	Il-12p40		[124]
	IFN- $\gamma$		
NSCLC	TGF- $\beta$	Decreased DCS concentration	[53]
Chemokines			
Oral, breast, melanoma	CXCR3	Tumor-LEC chemotaxis	[100]
	CXCR4-CXCL12	Tumor-LEC chemotaxis	[126]
Melanoma	CCR8-CCL1	Tumor-LEC chemotaxis	[125]
Gastric, colorectal, breast cancer	CCR7-CCL19/MIP-3 $\beta$	Tumor-LEC chemotaxis	[127]
	CCR7-CCL21/6Ckine	Tumor-LEC chemotaxis	[128]

SLN before cancer cell arrival [5]. Chemokine receptors such as CXCR3 and CXCR4 have been shown to be upregulated in animal models of different types of cancer and have been strongly associated with SLN metastasis [100]. Das et al. found that CCL1 protein was detected in the lymph node and lymphatic sinuses, and CCR8, the cognate receptor for CCL1, was significantly upregulated on the cell surface of human melanoma cells, providing a molecular basis for how CCL1 promotes cancer cell invasion into the SLN [125]. Several paired ligand-receptor pathways, including CXCL12-CXCR4, CCL19-CCR7 and CCL21-CCR7, also effectively promote the lymph node invasion of cancer cells [126–128]. These studies imply that the cytokine/chemokine milieu plays a crucial role in establishing a pre-metastatic microenvironment in the SLN.

#### Extracellular vesicles

Extracellular vesicles (EVs) are double layer lipid-containing vesicles that are naturally released from most types of cells. EVs can be divided into exosomes, microvesicles and apoptotic bodies based on their size and synthesis route [129]. These vesicles play important roles in cell–cell communication; they deliver different bioactive molecules from secreting cells to recipient cells to modulate the behaviors of recipient cells under various physiological and pathological conditions. Recent studies have highlighted the mechanisms of exosome-mediated processes in the preparation of pre-metastatic niches for lymphatic metastasis [130–132]. Tumor-secreted EVs can rebuild the surrounding matrix and reprogram the microenvironment in the lymph node to establish a beachhead for cancer cell spreading (Table. 2) [133–135]. For example, laminin 332, a large ECM protein complex, was found to be increased in the exocrine bodies of tumor tissues of patients with oral cancer with positive lymph node

**Table 2** SLN reprogramming with different cargo in EVs

Cancer type	Cargo in EVs	Effect	Refs.
OSCC	Lamin-332	Lymphangiogenesis	[136]
Colorectal cancer	IRF-2	Ingested by macrophage/induce VEGF-C expression	[137]
Breast cancer	Palmitoylated protein	NF $\kappa$ B activation via TLR-2 on macrophages	[139]
MDCK (normal kidney)	PDPN	Lymphangiogenesis	[140]
Melanoma, lung cancer, colorectal cancer	EGFR	Angiogenesis	[141]
Cervical cancer	miR221-3p	Induce LYVE-1 expression	[142]
Melanoma	PEDF	Recruitment of MDSCs/activation of TAM	[132, 143]
Melanoma	miR-9	Regulate angiogenesis via JAK-STAT	[144]

metastasis. Depletion of this protein potently suppresses EV-mediated LEC migration and lymphangiogenesis in the SLN [136]. Interferon regulatory factor 2 detected in plasma EVs is taken up by F4/80<sup>+</sup> macrophages to induce the release of VEGF-C to promote lymphangiogenesis and lymphatic network remodeling of the SLN in patients with colorectal cancer [137]. CD169<sup>+</sup> macrophages captured microvesicles derived from B16-F10 melanoma cells to trigger microenvironment reprogramming after their entry into the SLN [138]. Enhanced IL-6 expression in macrophages induced by breast cancer-released EV suppresses immune response and promotes cancer metastasis in a xenograft mouse model [139]. In addition, EV containing PDPN promotes tube formation in cultured LECs [140]. Moreover, exosomes derived from cancer cells can transmit EGFR to endothelial cells and LECs to stimulate angiogenesis and lymphangiogenesis [141]. In addition to proteins, nucleic acids in EVs have been found to be involved in lymph node metastasis. In cervical squamous cell carcinoma, miR-221-3p expression correlates with LYVE-1 expression and lymph node metastasis [142]. Cancer cell-derived miR-containing EVs enhance the recruitment of MDSC and the activation of TAM to generate an immunosuppressive environment, resulting in increased cancer cell metastasis [132, 143]. Melanoma-released exosomes delivered miR-9 to endothelial cells to activate Janus kinase 2-signal transducer and activator of transcription 3 signaling in endothelial cells to trigger angiogenesis [144]. These results suggest that EVs are critical mediators for cancer cells to remodel the SLN microenvironment.

### Metabolites

The lymph node is a lipid-rich organ because the lymph draining into it contains many fatty acids [44, 145]. Lee et al. reported that cancer cells undergo a metabolic shift toward fatty acid oxidation (FAO) via selective activation of a transcriptional coactivator, yes-associated protein (YAP), to promote lymph node metastasis [145]. FAO also plays a crucial role in controlling lymphangiogenesis [146]. Furthermore, FAO affects multiple cell types in the SLN. For instance, a recent study revealed the importance of FAO upregulation through STAT3 activation in inhibiting CD8<sup>+</sup> T-cell and in promoting obesity-associated breast tumorigenesis and metastasis [147]. The transcription factor Foxp3 increased the expression of acetyl-CoA synthetase and carnitine palmitoyltransferase 1A in Tregs, suggesting that Tregs prefer to use fatty acids as a major energy source; this process may contribute to functional reprogramming in the SLN [148]. Our study also demonstrated that bile acid metabolism and fatty acid metabolism are the hallmark pathways upregulated in the SLN. We found that overexpression of

fumarylacetoacetate hydrolase (FAH) in FRCs induced by breast cancer cells significantly increased mitochondrial OXPHOS levels and ATP production. FAH catalyzes the hydrolysis of 4-fumarylacetoacetate to acetoacetate and fumarate, which can be metabolized in the tricarboxylic acid cycle or used for biosynthetic purposes. Co-culture of immune cells isolated from mice with FAH-overexpressing FRCs inhibited immune cell activation *in vitro*, suggesting that metabolic reprogramming in FRCs produces metabolites that generate a tumor-induced immunosuppressive niche in the SLN. The concept of “oncometabolites” has been well established by the finding that metabolites, such as fumarate, succinate, and 2-hydroxyglutarate, can induce epigenetic alteration, enhance cellular transformation and generate a favorable microenvironment for tumor progression [149].

### Extracellular matrix

The ECM encompasses many extracellular macromolecules, including collagen, proteins, and hydroxyapatite. It generates a three-dimensional network that serves as a structural and biochemical basis to support tissue integrity. Under physiological circumstances, FRC is the major source of ECM production in the lymph nodes. They produce distinct types of ECM to form specific conduits in the lymph nodes to transport bioactive molecules and to speed up the migration of immune cells within lymph node [119, 120, 150]. Recently, Wei et al. identified a specific type of periostin<sup>+</sup> cancer-associated fibroblast that may promote lymph node metastasis in oral cancer by disrupting lymphatic endothelial barriers via the integrin-focal adhesion kinase/Src-VE-cadherin signaling pathway [151]. Li et al. elucidated that zinc finger protein 139 regulated annexin A proteins to accelerate lymph node metastasis in gastric cancer [152]. High expression of connexin-43 and E-cadherin was also found in the metastatic lymph nodes of gastric cancer [153]. Furthermore, an increase in enzymes such as lysyl oxidase, membrane type-matrix metalloproteinase and tissue inhibitor of matrix metalloproteinase 1 were associated with ECM degradation in the metastatic lymph nodes of patients with oral cancer [154]. In contrast, high levels of fibronectin, tenascin-C, and osteopontin in tumor stroma have been shown to be associated with lymph node metastasis [155, 156]. In summary, these studies indicate the importance of ECM remodeling in promoting lymph node metastasis.

### Therapeutic implication in targeting SLN

Because SLN involvement seems to be an initial and critical step for tumor metastasis, it could be a suitable target for therapeutic intervention by delivery of cytotoxic drugs or activation of immune response. Three strategies

are discussed herein. First, reversal of metabolic alteration. As previously mentioned, metabolic changes in various cell types in the SLN are found to be important for the creation of a pre-metastatic microenvironment. Therefore, inhibition or restoration of the metabolic switch is a considerable approach. For example, stromal cells in the SLN preferentially utilize lipids as a major energy source and undergo FAO for ATP production. Suppressing FAO reaction locally in the SLN may reverse the microenvironment to normal immune active status to reduce cancer cell arrival and invasion. Second, anti-cancer drugs can be directly delivered to SLN or regional lymph nodes to kill cancer cells. The main challenge of this approach is the development of lymph node-tropic nanoparticle. The size and characteristic of nanoparticle are key determinants for specific lymph node targeting. It has been shown that nanoparticles ranging 5 to 50 nm in size are favorable for uptake by lymphatic capillary and vessel, thus promoting SLN entry [157, 158]. Magnetic drug-conjugated nanoparticle can be concentrated in regional lymph nodes using an external magnet [159, 160]. The design of a drug-delivery particle with high SLN tropism is an important issue in nanomedicine. Third, enhancement of anti-cancer immunity in the SLN by immune modulators and cancer vaccines. Immunotherapy has become one of the mainstreams in cancer treatment. However, only a small proportion of patients with cancer benefit from immune checkpoint inhibitors. Recent studies demonstrated that SLN and tumor-draining lymph nodes are potential targets for re-enforcing immune responses. Sasso et al. developed lymphangiogenic potentiation of immunotherapy by injecting of VEGF-C-overexpressing and adjuvant-covering killed cancer cells to elicit T cell activation at the injection sites and draining lymph nodes [161]. Francis et al. also showed that locoregional delivery of immune checkpoint inhibitors in lymph nodes triggered enhanced cancer immunotherapy in the animal model of melanoma [162]. Intranodal injection of tumor-specific antigens increases DC presentation and promotes the efficacy of cancer vaccines. These results suggest that targeting SLN or tumor-draining lymph nodes could be a new direction for the development of anti-cancer drugs.

## Conclusions

In a landing battle, the establishment of a beachhead is the most critical step in winning the war. To successfully disseminate to distant organs, cancer cells send many outpost troops to establish a beachhead in the SLN for further metastasis. These outpost troops could be macromolecules such as EVs, growth factors, and ECM proteins or small molecules such as miRNA

and metabolites. These secreted factors remodel all cell types, including immune cells, LEC, BEC, and fibroblasts, in the SLN and change the foe to friend to create a favorable microenvironment for cancer cell invasion. Our understanding of SLN reprogramming has vastly improved in the past two decades. In the near future, we can expect the application of lymph node-targeted drugs and vaccines for cancer treatment.

## Abbreviations

BEC: Blood endothelial cell; CAF: Cancer associated fibroblast; CCL21: C-C motif Chemokine ligand 21; CCR: CC-chemokine receptor; COX: Cyclooxygenase; Cox7c: Cytochrome c oxidase subunit 7c; CPT1A: Carnitine palmitoyl-transferase 1A; CTLA4: Cytotoxic T-lymphocyte-associated protein 4; CXCR: C-X-C motif chemokine receptor; 2-HG: 2-hydroxyglutarate; DC: Dendritic cell; DEG: Differentially expressed gene; ECM: Extracellular matrix; ER: Estrogen receptor; EV: Extracellular vesicle; FAH: Fumarylacetoacetate hydroxylase; FAK: Focal adhesion kinase; FAO: Fatty acid oxidation; FDC: Follicular dendritic cell; Foxp3: Forkhead box P3; FRC: Fibroblastic reticular cell; GM-CSF: Granulocyte macrophage-colony stimulating factor; HEV: High endothelial venules; IDO: Indoleamine 2,3-dioxygenase; IFN: Interferon; IL: Interleukin; LEC: Lymphatic endothelial cell; LOX: Lysyl oxidase; LTO: Lymphoid tissue organizer; LYVE-1: Lymphatic vessel endothelial hyaluronan receptor 1; M-CSF: Macrophage colony-stimulating factor; MDSC: Myeloid-derived suppressor cells; MEF2C: Myocyte-specific enhancer factor 2C; MHC: Major histocompatibility complex; Ndufv2: Ubiquinone oxidoreductase core subunit v2; OCR: Oxygen consumption rate; OXPHOS: Oxidative phosphorylation; PDGF: Platelet-derived growth factor; PDPN: Podoplanin; PGC-1 $\alpha$ : Proliferator-activated receptor gamma coactivator 1 alpha; PGE2: Prostaglandin E2; PI3K: Phosphatidylinositol-3-kinase; PNA: Peripheral node addressin; Prdx5: Peroxiredoxin-5; Prox-1: Prospero homeobox protein 1; SDHD: Succinate dehydrogenase complex subunit D; SLN: Sentinel lymph nodes; STAT: Signal transducer and activator of transcription; TAM: Tumor-associated macrophage; TFAM: Transcription factor A, mitochondrial; TGF- $\beta$ : Transforming Growth Factor Beta; Treg: Regulatory T cell; Uqcrl1: Ubiquinol-cytochrome c reductase, complex III subunit XI; VEGF: Vascular endothelial growth factor; VEGFR-3: Vascular endothelial growth factor receptor 3; YAP: Yes-associated protein.

## Acknowledgements

We thank Dr. Mei-Ren Pan and Dr. Shu-Yi Lin for value suggestion in the preparation of the manuscript.

## Author contributions

YLL and WCH wrote the manuscript. Both authors read and approved the final manuscript.

## Funding

This study was supported by the grants 110-2320-B-400-016 from Ministry of Science and Technology, Taiwan and CA-111-PP-17 from Ministry of Health and Welfare, Taiwan to Wen-Chun Hung.

## Availability of data and materials

Not applicable.

## Declarations

### Ethics approval and consent to participate

Not applicable.

### Consent for publication

Not applicable.

### Competing interests

All authors declare no conflict of interest.

**Author details**

<sup>1</sup>National Institute of Cancer Research, National Health Research Institutes, Tainan 704, Taiwan. <sup>2</sup>School of Pharmacy, College of Pharmacy, Kaohsiung Medical University, Kaohsiung 807, Taiwan.

Received: 27 April 2022 Accepted: 15 October 2022

Published online: 20 October 2022

**References**

- Oliver G, Kipnis J, Randolph GJ, Harvey NL. The lymphatic vasculature in the 21st century: Novel functional roles in homeostasis and disease. *Cell*. 2020;182(2):270–96.
- Ruddle NH. Lymphatic vessels and tertiary lymphoid organs. *J Clin Invest*. 2014;124(3):953–9.
- Mohammed RAA, Martin SG, Gill MS, Green AR, Paish EC, Ellis IO. Improved methods of detection of lymphovascular invasion demonstrate that it is the predominant method of vascular invasion in breast cancer and has important clinical consequences. *Am J Surg Pathol*. 2007;31(12):1825–33.
- Chao NQ, Berghuis B, Tsarfaty G, Bruch BM, Kort EJ, Ditlev J, et al. Preparing the "soil": The primary tumor induces vasculature reorganization in the sentinel lymph node before the arrival of metastatic cancer cells. *Cancer Res*. 2006;66(21):10365–76.
- Jones D. Parallels of resistance between angiogenesis and lymphangiogenesis inhibition in cancer therapy. *Cell*. 2020;9(3):762.
- Ndiaye PD, Pagès G. VEGF-C and lymphatic vessels: a double-edged sword in tumor development and metastasis. *Med Sci (Paris)*. 2019;35(2):132–7.
- Künnapuu J, Bokharaie H, Jeltsch M. Proteolytic cleavages in the VEGF family: Generating diversity among angiogenic VEGFs, essential for the activation of lymphangiogenic VEGFs. *Biology (Basel)*. 2021;10(2):167.
- Sha M, Jeong S, Chen XS, Tong Y, Cao J, Sun HY, et al. Expression of VEGFR-3 in intrahepatic cholangiocarcinoma correlates with unfavorable prognosis through lymphangiogenesis. *Int J Biol Sci*. 2018;14(10):1333–42.
- Kwon S, Agollah GD, Wu G, Chan W, Sevick-Muraca EM. Direct visualization of changes of lymphatic function and drainage pathways in lymph node metastasis of B16F10 melanoma using near-infrared fluorescence imaging. *Biomed Opt Express*. 2013;4(6):967–77.
- Morein D, Erlichman N, Ben-Baruch A. Beyond cell motility: the expanding roles of chemokines and their receptors in malignancy. *Front Immunol*. 2020;11:952.
- Poeta VM, Massara M, Capucetti A, Bonecchi R. Chemokines and chemokine receptors: New targets for cancer immunotherapy. *Front Immunol*. 2019;10:379.
- Skobe M, Hawighorst T, Jackson DG, Prevo R, Janes L, Velasco P, et al. Induction of tumor lymphangiogenesis by VEGF-C promotes breast cancer metastasis. *Nat Med*. 2001;7(2):192–8.
- Mandriota SJ, Jussila L, Jeltsch M, Compagni A, Baetens D, Prevo R, et al. Vascular endothelial growth factor-C-mediated lymphangiogenesis promotes tumour metastasis. *EMBO J*. 2001;20(4):672–82.
- Hirakawa S, Kodama S, Kunstfeld R, Kajiya K, Brown LF, Detmar M. VEGF-A induces tumor and sentinel lymph node lymphangiogenesis and promotes lymphatic metastasis. *J Exp Med*. 2005;201(7):1089–99.
- Björndahl MA, Cao R, Burton JB, Brakenhielm E, Religa P, Galter D, et al. Vascular endothelial growth factor- $\alpha$  promotes peritumoral lymphangiogenesis and lymphatic metastasis. *Cancer Res*. 2005;65(20):9261–8.
- Thelen A, Scholz A, Benckert C, Marschall ZV, Schröder M, Wiedenmann B, et al. VEGF-D promotes tumor growth and lymphatic spread in a mouse model of hepatocellular carcinoma. *Int J Cancer*. 2008;122(11):2471–81.
- Cao R, Björndahl MA, Religa P, Clasper S, Garvin S, Galter D, et al. PDGF-BB induces intratumoral lymphangiogenesis and promotes lymphatic metastasis. *Cancer Cell*. 2004;6(4):333–45.
- Nguyen-Ngoc KV, Cheung KJ, Brenot A, Shamir ER, Gray RS, Hines WC, et al. ECM microenvironment regulates collective migration and local dissemination in normal and malignant mammary epithelium. *Proc Natl Acad Sci USA*. 2012;109(39):E2595–604.
- Moehrl M, Schippert W, Rassner G, Garbe C, Breuninger H. Micrometastasis of a sentinel lymph node in cutaneous melanoma is a significant prognostic factor for disease-free survival, distant-metastasis-free survival, and overall survival. *Dermatol Surg*. 2004;30(10):1319–28.
- Leiter U, Buettner PG, Bohnenberger K, Eigentler T, Meier F, Moehrl M, et al. Sentinel lymph node dissection in primary melanoma reduces subsequent regional lymph node metastasis as well as distant metastasis after nodal involvement. *Ann Surg Oncol*. 2010;17(1):129–37.
- Grigoriadis A, Gazinska P, Pai T, Irhsad S, Wu Y, Millis R, et al. Histological scoring of immune and stromal features in breast and axillary lymph nodes is prognostic for distant metastasis in lymph node-positive breast cancers. *J Pathol Clin Res*. 2018;4(1):39–54.
- Zhu Y, Lin J, Yan Y, Zheng K, Zhang H, Wu K, et al. Delphian lymph node metastasis is a novel indicator of tumor aggressiveness and poor prognosis in papillary thyroid cancer. *J Surg Oncol*. 2021;123(7):1521–8.
- Naxerova K, Reiter JG, Brachtel E, Lennerz JK, van de Wetering M, Rowan A, et al. Origins of lymphatic and distant metastases in human colorectal cancer. *Science*. 2017;357(6346):55–60.
- Venet D, Fimereli D, Rothé F, Boeckx B, Maetens M, Majaj S, et al. Phylogenetic reconstruction of breast cancer reveals two routes of metastatic dissemination associated with distinct clinical outcome. *EBioMedicine*. 2020;56: 102793.
- Reticker-Flynn NE, Zhang W, Belk JA, Basto PA, Escalante NK, Pilarowski OGW, et al. Lymph node colonization induces tumor-immune tolerance to promote distant metastasis. *Cell*. 2022;185(11):1924–42.
- Faries MB, Thompson JF, Cochran AJ, Andtbacka RH, Mozzillo N, Zager JS, et al. Completion dissection or observation for sentinel-node metastasis in melanoma. *N Engl J Med*. 2017;376:2211–22.
- Bozec A, Dassonville O, Chamorey E, Poissonnet G, Sudaka A, Peyrottes I, et al. Clinical impact of cervical lymph node involvement and central neck dissection in patients with papillary thyroid carcinoma: a retrospective analysis of 368 cases. *Eur Arch Otorhinolaryngol*. 2011;268(8):1205–12.
- Giuliano AE, Ballman KV, McCall L, Beitsch PD, Brennan MB, Kelemen PR, et al. Effect of axillary dissection vs no axillary dissection on 10-year overall survival among women with invasive breast cancer and sentinel node metastasis: the ACOSOG Z0011 (Alliance) randomized clinical trial. *JAMA*. 2017;318(10):918–26.
- Fisher B, Jeong JH, Anderson S, Bryant J, Fisher ER, Wolmark N. Twenty-five-year follow-up of a randomized trial comparing radical mastectomy, total mastectomy, and total mastectomy followed by irradiation. *N Engl J Med*. 2002;347(8):567–75.
- Kuo YT, Tsai WS, Hung HY, Hsieh PS, Chiang SF, Lai CC, et al. Prognostic value of regional lymph node involvement in patients with metastatic colorectal cancer: palliative versus curative resection. *World J Surg Oncol*. 2021;19(1):150.
- Ballester M, Dubernard G, Lécure F, Heitz D, Mathevet P, Marret H, et al. Detection rate and diagnostic accuracy of sentinel-node biopsy in early stage endometrial cancer: a prospective multicentre study (SENTI-ENDO). *Lancet Oncol*. 2011;12(5):469–76.
- Joshi NP, Broughman JR. Postoperative management of salivary gland tumors. *Curr Treat Options Oncol*. 2021;22(3):23.
- Fonseca IB, Lindote MVN, Monteiro MR, Filho ED, Pinto CAL, Jafelicci AS, et al. Sentinel node status is the most important prognostic information for clinical stage IIB and IIC melanoma patients. *Ann Surg Oncol*. 2020;27(11):4133–40.
- Tropea S, Fiore PD, Maurichi A, Patuzzo R, Santinami M, Ribero S, et al. The role of sentinel node tumor burden in modeling the prognosis of melanoma patients with positive sentinel node biopsy: an Italian melanoma intergroup study (N = 2,086). *BMC Cancer*. 2022;22(1):610.
- Straker RJ 3rd, Sharon CE, Chu EY, Miura JT, Ming ME, Karakousis GC. Sentinel lymph node biopsy in patients with clinical stage IIB/C cutaneous melanoma: a national cohort study. *J Am Acad Dermatol*. 2022;87(4):754–60.
- Saha S, Philimon B, Efeson M, Helina A, Elgamal M, Kiya G, et al. The role of sentinel lymph node mapping in colon cancer: detection of micrometastasis, effect on survival, and driver of a paradigm shift in extent of colon resection. *Clin Exp Metastasis*. 2022;39(1):109–15.
- Honselmann KC, Pergolini I, Fernandez-Del Castillo C, Deshpande V, Ting D, Taylor MS, et al. Timing but not patterns of recurrence is



- different between node-negative and node-positive resected pancreatic cancer. *Ann Surg*. 2020;272(2):357–65.
38. Wang F, Ge X, Wang Z, Weng Y, Yin R, You Q. Clinical significance and prognosis of supraclavicular lymph node metastasis in patients with thoracic esophageal cancer. *Ann Transl Med*. 2020;8(4):90.
  39. Chang GJ, Rodriguez-Bigas MA, Skibber JM, Moyer VA. Lymph node evaluation and survival after curative resection of colon cancer: systematic review. *J Natl Cancer Inst*. 2007;99(6):433–41.
  40. Pastushenko I, Van den Eynden GG, Arregui SV, Torres LP, Alegret RA, Querol I, et al. Increased angiogenesis and lymphangiogenesis in metastatic sentinel lymph nodes is associated with nonsentinel lymph node involvement and distant metastasis in patients with melanoma. *Am J Dermatopathol*. 2016;38(5):338–46.
  41. Blenman KRM, He TF, Frankel PH, Ruel NH, Schwartz EJ, Krag DN, et al. Sentinel lymph node B cells can predict disease-free survival in breast cancer patients. *NPJ Breast Cancer*. 2018;4:28.
  42. Singh V. *Textbook of Anatomy: Head, Neck, and Brain*. 2nd ed. ELSEVIER Ltd.; 2017.
  43. Warwick R, Williams PL. *Angiology (Chapter 6). Gray's anatomy*, 35 edn, Longman, Ltd.; 1973.
  44. Kawada K, Taketo MM. Significance and mechanism of lymph node metastasis in cancer progression. *Cancer Res*. 2011;71(4):1214–8.
  45. Edge SB, Compton CC. The American Joint Committee on Cancer: The 7th edition of the AJCC cancer staging manual and the future of TNM. *Ann Surg Oncol*. 2010;17(6):1471–4.
  46. Nathanson SD, Shah R, Rosso K. Sentinel lymph node metastases in cancer: causes, detection and their role in disease progression. *Semin Cell Dev Biol*. 2015;38:106–16.
  47. Harrell MI, Iritani BM, Ruddell A. Tumor-induced sentinel lymph node lymphangiogenesis and increased lymph flow precede melanoma metastasis. *Am J Pathol*. 2007;170(2):774–86.
  48. Chung MK, Do IG, Jung E, Son YI, Jeong HS, Baek CH. Lymphatic vessels and high endothelial venules are increased in the sentinel lymph nodes of patients with oral squamous cell carcinoma before the arrival of tumor cells. *Ann Surg Oncol*. 2012;19(5):1595–601.
  49. Cheng R, Billet S, Liu C, Haldar S, Choudhury D, Tripathi M, et al. Periodontal inflammation recruits distant metastatic breast cancer cells by increasing myeloid-derived suppressor cells. *Oncogene*. 2020;39(7):1543–56.
  50. Gibert-Ramos A, López C, Bosch R, Fontoura L, Bueno G, García-Rojo M, et al. Immune response profile of primary tumour, sentinel and non-sentinel axillary lymph nodes related to metastasis in breast cancer: an immunohistochemical point of view. *Histochem Cell Biol*. 2019;152(3):177–93.
  51. Ito M, Minamiya Y, Kawai H, Saito S, Saito H, Nakagawa T, et al. Tumor-derived TGFβ-1 induces dendritic cell apoptosis in the sentinel lymph node. *J Immunol*. 2006;176(9):5637–43.
  52. Kaplan RN, Riba RD, Zacharoulis S, Bramley AH, Vincent L, Costa C, et al. VEGFR1-positive haematopoietic bone marrow progenitors initiate the pre-metastatic niche. *Nature*. 2005;438(7069):820–7.
  53. Lund AW, Duraes FV, Hirosue S, Raghavan VR, Nembrini C, Thomas SN, et al. VEGF-C promotes immune tolerance in B16 melanomas and cross-presentation of tumor antigen by lymph node lymphatics. *Cell Rep*. 2012;1(3):191–9.
  54. Munn DH, Sharma MD, Hou D, Baban B, Lee JR, Antonia SJ, et al. Expression of indoleamine 2,3-dioxygenase by plasmacytoid dendritic cells in tumor-draining lymph nodes. *J Clin Invest*. 2004;114(2):280–90.
  55. Sharma MD, Baban B, Chandler P, Hou DY, Singh N, Yagita H, et al. Plasmacytoid dendritic cells from mouse tumor-draining lymph nodes directly activate mature Tregs via indoleamine 2,3-dioxygenase. *J Clin Invest*. 2007;117(9):2570–82.
  56. Sakakura K, Chikamatsu K, Sakurai T, Takahashi K, Murata T, Oriuchi N, et al. Infiltration of dendritic cells and NK cells into the sentinel lymph node in oral cavity cancer. *Oral Oncol*. 2005;41(1):89–96.
  57. Farzad Z, Cochran AJ, McBride WH, Gray JD, Wong V, Morton DL. Lymphocyte subset alterations in nodes regional to human melanoma. *Cancer Res*. 1990;50(12):3585–8.
  58. Piersiala K, Silva PFN, Hjalmarsson E, Kolev A, Kågedal A, Starkhammar M, et al. CD4+ and CD8+ T cells in sentinel nodes exhibit distinct pattern of PD-1, CD69, and HLA-DR expression compared to tumor tissue in oral squamous cell carcinoma. *Cancer Sci*. 2021;112(3):1048–59.
  59. Cesana GC, DeRaffele G, Cohen S, Moroziewicz D, Mitcham J, Stoutenburg J, et al. Characterization of CD4+CD25+ regulatory T cells in patients treated with high-dose interleukin-2 for metastatic melanoma or renal cell carcinoma. *J Clin Oncol*. 2006;24(7):1169–77.
  60. Viguier M, Lemaitre F, Verola O, Cho MS, Gorochov G, Dubertret L, et al. Foxp3 expressing CD4+CD25(high) regulatory T cells are overrepresented in human metastatic melanoma lymph nodes and inhibit the function of infiltrating T cells. *J Immunol*. 2004;173(2):1444–53.
  61. Faghieh Z, Erfani N, Haghshenas MR, Safaei A, Talei AR, Ghaderi A. Immune profiles of CD4+ lymphocyte subsets in breast cancer tumor draining lymph nodes. *Immunol Lett*. 2014;158(1–2):57–65.
  62. Schule JM, Bergkvist L, Hakansson L, Gustafsson B, Hakansson A. CD28 expression in sentinel node biopsies from breast cancer patients in comparison with CD3-zeta chain expression. *J Transl Med*. 2004;2(1):45.
  63. Ruddell A, Harrell MI, Furuya M, Kirschbaum SB, Iritani BM. B lymphocytes promote lymphogenous metastasis of lymphoma and melanoma. *Neoplasia*. 2011;13(8):748–57.
  64. Zirakzadeh AA, Marits P, Sherif A, Winqvist O. Multiplex B cell characterization in blood, lymph nodes, and tumors from patients with malignancies. *J Immunol*. 2013;190(11):5847–55.
  65. Li Q, Lao X, Pan Q, Ning N, Yet J, Xu Y, et al. Adoptive transfer of tumor reactive B cells confers host T-cell immunity and tumor regression. *Clin Cancer Res*. 2011;17(15):4987–95.
  66. Li YL, Chen CH, Chen JY, Lai YS, Wang SC, Jiang SS, et al. Single-cell analysis reveals immune modulation and metabolic switch in tumor-draining lymph nodes. *Oncoimmunology*. 2020;9(1):1830513.
  67. Qian Y, Zhai E, Chen S, Liu Y, Ma Y, Chen J, et al. Single-cell RNA-seq dissecting heterogeneity of tumor cells and comprehensive dynamics in tumor microenvironment during lymph nodes metastasis in gastric cancer. *Int J Cancer*. 2022;151(8):1367–81.
  68. Chen RY, Fan YM, Zhang Q, Liu S, Li Q, Ke GL, et al. Estradiol inhibits Th17 cell differentiation through inhibition of RORγT transcription by recruiting the ERα/REα complex to estrogen response elements of the RORγT promoter. *J Immunol*. 2015;194(8):4019–28.
  69. Kim DH, Park HJ, Park HS, Lee JU, Ko C, Gye MC, et al. Estrogen receptor alpha in T cells suppresses follicular helper T cell responses and prevents autoimmunity. *Exp Mol Med*. 2019;51(4):1–9.
  70. Yang C, Lee H, Pal S, Jove V, Deng J, Zhang W, et al. B cells promote tumor progression via STAT3 regulated-angiogenesis. *PLoS ONE*. 2013;8(5): e64159.
  71. Wilker PR, Kohyama M, Sandau MM, Albring JC, Nakagawa O, Schwarz JJ, et al. Transcription factor Mef2c is required for B cell proliferation and survival after antigen receptor stimulation. *Nat Immunol*. 2008;9(6):603–12.
  72. Herglotz J, Unrau L, Hauschildt F, Fischer M, Kriebitzsch N, Alawi M, et al. Essential control of early B-cell development by Mef2 transcription factors. *Blood*. 2016;127(5):572–81.
  73. Wang W, Org T, Montel-Hagen A, Pioli PD, Duan D, Israely E, et al. MEF2C protects bone marrow B-lymphoid progenitors during stress haematopoiesis. *Nat Commun*. 2016;7:12376.
  74. Tamayo E, Alvarez P, Merino R. TGFβ family members as regulators of B Cell development and functional implications for autoimmunity. *Int J Mol Sci*. 2018;19(12):3928.
  75. Kitamura T, Qian BZ, Pollard JW. Immune cell promotion of metastasis. *Nat Rev Immunol*. 2015;15(2):73–86.
  76. Schaller J, Agudo J. Metastatic colonization: Escaping immune surveillance. *Cancers (Basel)*. 2020;12(11):3385.
  77. Pan PY, Ma G, Weber KJ, Ozao-Choy J, Wang G, Yin B, et al. Immune stimulatory receptor CD40 is required for T-cell suppression and T regulatory cell activation mediated by myeloid-derived suppressor cells in cancer. *Cancer Res*. 2010;70(11):99–108.
  78. Huang B, Pan PY, Li Q, Sato AI, Levy DE, Bromberg J, et al. Gr-1+CD115+ immature myeloid suppressor cells mediate the development of tumor-induced T regulatory cells and T-cell anergy in tumor-bearing host. *Cancer Res*. 2006;66(2):1123–31.
  79. Serafini P, Mgebroff S, Noonan K, Borrello I. Myeloid-derived suppressor cells promote cross-tolerance in B-cell lymphoma by expanding regulatory T cells. *Cancer Res*. 2008;68(13):5439–49.
  80. Bian Z, Abdelaal AM, Shi L, Liang H, Xiong L, Kidder K, et al. Arginase-1 is neither constitutively expressed in nor required for myeloid-derived

- suppressor cell-mediated inhibition of T-cell proliferation. *Eur J Immunol*. 2018;48(6):1046–58.
81. Grzywa TM, Sosnowska A, Matryba P, Rydzynska Z, Jasinski M, Nowis D, et al. Myeloid cell-derived arginase in cancer immune response. *Front Immunol*. 2020;11:938.
  82. Ohl K, Tenbrock K. Reactive oxygen species as regulators of MDSC-mediated immune suppression. *Front Immunol*. 2018;9:2499.
  83. Feng S, Cheng X, Zhang L, Lu X, Chaudhary S, Teng R, et al. Myeloid-derived suppressor cells inhibit T cell activation through nitrating LCK in mouse cancers. *Proc Natl Acad Sci USA*. 2018;115(40):10094–9.
  84. Molon B, Ugel S, Del P, Soldani C, Zilio S, Avella D, et al. Chemokine nitration prevents intratumoral infiltration of antigen-specific T cells. *J Exp Med*. 2011;208(10):1949–62.
  85. Ku AW, Muhitch JB, Powers CA, Diehl M, Kim M, Fisher DT, et al. Tumor-induced MDSC act via remote control to inhibit L-selectin-dependent adaptive immunity in lymph nodes. *Elife*. 2016;5: e17375.
  86. Cochran AJ, Huang RR, Su A, Itakura E, Wen DR. Is sentinel node susceptibility to metastases related to nodal immune modulation? *Cancer J*. 2015;21(1):39–46.
  87. Yu J, Du W, Yan F, Wang Y, Li H, Cao S, et al. Myeloid-derived suppressor cells suppress antitumor immune responses through IDO expression and correlate with lymph node metastasis in patients with breast cancer. *J Immunol*. 2013;190(7):3783–97.
  88. Go Y, Tanaka H, Tokumoto M, Sakurai K, Toyokawa T, Kubo N, et al. Tumor-associated macrophages extend along lymphatic flow in the pre-metastatic lymph nodes of human gastric cancer. *Ann Surg Oncol*. 2016;23(Suppl 2):S230–5.
  89. Saito Y, Ohnishi K, Miyashita A, Nakahara S, Fujiwara Y, Horlad H, et al. Prognostic significance of CD169+ lymph node sinus macrophages in patients with malignant melanoma. *Cancer Immunol Res*. 2015;3(12):1356–63.
  90. Shiota T, Miyasato Y, Ohnishi K, Yamamoto-Ibusuki M, Yamamoto Y, Iwase H, et al. The clinical significance of CD169-positive lymph node macrophage in patients with breast cancer. *PLoS ONE*. 2016;11(11): e0166680.
  91. Stromvall K, Sundkvist K, Ljungberg B, Halin Bergstrom S, Bergh A. Reduced number of CD169(+) macrophages in pre-metastatic regional lymph nodes is associated with subsequent metastatic disease in an animal model and with poor outcome in prostate cancer patients. *Prostate*. 2017;77(15):1468–77.
  92. Aspelund A, Robciuc MR, Karaman S, Makinen T, Alitalo K. Lymphatic system in cardiovascular medicine. *Circ Res*. 2016;118:515–30.
  93. Brulois K, Rajaraman A, Szade A, Nordling S, Bogoslawski A, Dermadi D, et al. A molecular map of murine lymph node blood vascular endothelium at single cell resolution. *Nat Commun*. 2020;11(1):3798.
  94. Lee M, Kiefel H, Lajevic MD, Macauley MS, Kawashima H, O'Hara E, et al. Transcriptional programs of lymphoid tissue capillary and high endothelium reveal control mechanisms for lymphocyte homing. *Nat Immunol*. 2014;15(10):982–95.
  95. Mionnet C, Sanos SL, Mondor I, Jorquera A, Laugier JP, Germain RN, et al. High endothelial venules as traffic control points maintaining lymphocyte population homeostasis in lymph nodes. *Blood*. 2011;118(23):6115–22.
  96. Miyasaka M, Tanaka T. Lymphocyte trafficking across high endothelial venules: dogmas and enigmas. *Nat Rev Immunol*. 2004;4(5):360–70.
  97. Padera TP, Meijer EF, Munn LL. The lymphatic system in disease processes and cancer progression. *Annu Rev Biomed Eng*. 2016;18:125–58.
  98. Lee SY, Qian CN, Ooi AS, Chen P, Wong HM, Myint SS, et al. Changes in specialized blood vessels in lymph nodes and their role in cancer metastasis. *J Transl Med*. 2012;10:206.
  99. McDonald D. Angiogenesis and vascular remodeling in inflammation and cancer: Biology and architecture of the vasculature. *Angiogenesis*. 2008. [https://doi.org/10.1007/978-0-387-71518-6\\_2](https://doi.org/10.1007/978-0-387-71518-6_2).
  100. Stacker SA, Williams SP, Karnezis T, Shayan R, Fox SB, Achen MG. Lymphangiogenesis and lymphatic vessel remodelling in cancer. *Nat Rev Cancer*. 2014;14(3):159–72.
  101. Zheng W, Aspelund A, Alitalo K. Lymphangiogenic factors, mechanisms, and applications. *J Clin Invest*. 2014;124(3):878–87.
  102. Arimoto S, Hasegawa T, Takeda D, Saito I, Amano R, Akashi M, et al. Lymphangiogenesis and lymph node metastasis in oral squamous cell carcinoma. *Anticancer Res*. 2018;38(11):6157–62.
  103. Bosisio D, Ronca R, Salvi V, Presta M, Sozzani S. Dendritic cells in inflammatory angiogenesis and lymphangiogenesis. *Curr Opin Immunol*. 2018;53:180–6.
  104. Fujimoto N, He Y, D'Addio M, Tacconi C, Detmar M, Dieterich LC. Single-cell mapping reveals new markers and functions of lymphatic endothelial cells in lymph nodes. *PLoS Biol*. 2020;18(4): e3000704.
  105. Pan Z, Lu X, Zhao J, Gao Q, Wang J. VEGF-C is positively associated with lymphangiogenesis and lymphatic metastasis in rectal cancer. *Int J Clin Exp Pathol*. 2018;11(3):1777–83.
  106. Garmy-Susini B, Avraamides CJ, Desgrosellier JS, Schmid MC, Foubert P, Ellices LG, et al. PI3Kalpha activates integrin alpha4beta1 to establish a metastatic niche in lymph nodes. *Proc Natl Acad Sci USA*. 2013;110(22):9042–7.
  107. Tutunea-Fatan E, Majumder M, Xin X, Lala PK. The role of CCL21/CCR7 chemokine axis in breast cancer-induced lymphangiogenesis. *Mol Cancer*. 2015;14:35.
  108. Karnezis T, Shayan R, Fox S, Achen MG, Stacker SA. The connection between lymphangiogenic signalling and prostaglandin biology: a missing link in the metastatic pathway. *Oncotarget*. 2012;3(8):893–906.
  109. Witte MH, Jones K, Wilting J, Dictor M, Selg M, McHale N, et al. Structure function relationships in the lymphatic system and implications for cancer biology. *Cancer Metastasis Rev*. 2006;25(2):159–84.
  110. Avery M, Nathanson SD, Hetzel FW. Lymph flow from murine footpad tumors before and after sublethal hyperthermia. *Radiat Res*. 1992;132(1):50–3.
  111. Cordeiro OG, Chypre M, Brouard N, Rauber S, Allouh F, Romera-Hernandez M, et al. Integrin-alpha 11b identifies murine lymph node lymphatic endothelial cells responsive to RANKL. *PLoS ONE*. 2016;11(3): e0151848.
  112. Rouhani SJ, Eccles JD, Tewalt EF, Engelhard VH. Regulation of T-cell tolerance by lymphatic endothelial cells. *J Clin Cell Immunol*. 2014;5:1000242.
  113. Jalkanen S, Salmi M. Lymphatic endothelial cells of the lymph node. *Nat Rev Immunol*. 2020;20(9):566–78.
  114. Card CM, Yu SS, Swartz MA. Emerging roles of lymphatic endothelium in regulating adaptive immunity. *J Clin Invest*. 2014;124(3):943–52.
  115. Cohen JN, Guidi CJ, Tewalt EF, Qiao H, Rouhani SJ, Ruddell A, et al. Lymph node-resident lymphatic endothelial cells mediate peripheral tolerance via Aire-independent direct antigen presentation. *J Exp Med*. 2010;207(4):681–8.
  116. Tewalt EF, Cohen JN, Rouhani SJ, Engelhard VH. Lymphatic endothelial cells -key players in regulation of tolerance and immunity. *Front Immunol*. 2012;3:305.
  117. Habenicht LM, Kirschbaum SB, Furuya M, Harrell MI, Ruddell A. Tumor regulation of lymph node lymphatic sinus growth and lymph flow in mice and in humans. *Yale J Biol Med*. 2017;90(3):403–15.
  118. Tewalt EF, Cohen JN, Rouhani SJ, Guidi CJ, Qiao H, Fahl SP, et al. Lymphatic endothelial cells induce tolerance via PD-L1 and lack of costimulation leading to high-level PD-1 expression on CD8 T cells. *Blood*. 2012;120(24):4772–82.
  119. Morgado FN, Silva AVA, Porrozzi R. Infectious diseases and the lymphoid extracellular matrix remodeling: A focus on conduit system. *Cells*. 2020;9(3):725.
  120. Martinez VG, Pankova V, Krasny L, Singh T, Makris S, White IJ, et al. Fibroblastic reticular cells control conduit matrix deposition during lymph node expansion. *Cell Rep*. 2019;29(9):2810–22.
  121. Riedel A, Shorthouse D, Haas L, Hall BA, Shields J. Tumor-induced stromal reprogramming drives lymph node transformation. *Nat Immunol*. 2016;17(9):1118–27.
  122. Botella-Estrada R, Dasi F, Ramos D, Nagore E, Herrero MJ, Gimenez J, et al. Cytokine expression and dendritic cell density in melanoma sentinel nodes. *Melanoma Res*. 2005;15(2):99–106.
  123. Leong SP, Peng M, Zhou YM, Vaquerano JE, Chang JW. Cytokine profiles of sentinel lymph nodes draining the primary melanoma. *Ann Surg Oncol*. 2002;9(1):82–7.
  124. Matsuura K, Yamaguchi Y, Ueno H, Osaki A, Arihiro K, Toge T. Maturation of dendritic cells and T-cell responses in sentinel lymph nodes from patients with breast carcinoma. *Cancer*. 2006;106(6):1227–36.
  125. Cera MR, Del Prete A, Vecchi A, Corada M, Martin-Padura J, Motoike T, et al. Increased DC trafficking to lymph nodes and contact hypersensitivity in junctional adhesion molecule-A-deficient mice. *J Clin Invest*. 2004;114(5):729–38.

126. Rusetska N, Kowalski K, Zalewski K, Zięba S, Bidziński M, Goryca K, et al. CXCR4/ACKR3/CXCL12 axis in the lymphatic metastasis of vulvar squamous cell carcinoma. *J Clin Pathol*. 2021;75(5):324–32.
127. Liu Q, Qiao M, Lohinai Z, Mao S, Pan Y, Wang Y, et al. CCL19 associates with lymph node metastasis and inferior prognosis in patients with small cell lung cancer. *Lung Cancer*. 2021;162:194–202.
128. Hayasaka H, Yoshida J, Kuroda Y, Nishiguchi A, Matsusaki M, Kishimoto K, et al. CXCL12 promotes CCR7 ligand-mediated breast cancer cell invasion and migration toward lymphatic vessels. *Cancer Sci*. 2022;113(4):1338–51.
129. Veziroglu EM, Mias GI. Characterizing extracellular vesicles and their diverse RNA contents. *Front Genet*. 2020;11:700.
130. Becker A, Thakur BK, Weiss JM, Kim HS, Peinado H, Lyden D. Extracellular vesicles in cancer: cell-to-cell mediators of metastasis. *Cancer Cell*. 2016;30(6):836–48.
131. Broggi MAS, Maillat L, Clement CC, Bordry N, Corthesy P, Auger A, et al. Tumor-associated factors are enriched in lymphatic exudate compared to plasma in metastatic melanoma patients. *J Exp Med*. 2019;216(5):1091–107.
132. Hood JL, San RS, Wickline SA. Exosomes released by melanoma cells prepare sentinel lymph nodes for tumor metastasis. *Cancer Res*. 2011;71(11):3792–801.
133. Nicolini A, Ferrari P, Biava PM. Exosomes and cell communication: from tumour-derived exosomes and their role in tumour progression to the use of exosomal cargo for cancer treatment. *Cancers (Basel)*. 2021;13(4):822.
134. Sleeman JP. The lymph node pre-metastatic niche. *J Mol Med (Berl)*. 2015;93(11):1173–84.
135. Wang CA, Tsai SJ. Regulation of lymphangiogenesis by extracellular vesicles in cancer metastasis. *Exp Biol Med (Maywood)*. 2021;246(19):2048–56.
136. Wang SH, Liou GG, Liu SH, Chang JS, Hsiao JR, Yen YC, et al. Laminin gamma2-enriched extracellular vesicles of oral squamous cell carcinoma cells enhance in vitro lymphangiogenesis via integrin alpha3-dependent uptake by lymphatic endothelial cells. *Int J Cancer*. 2019;144(11):2795–810.
137. Sun B, Zhou Y, Fang Y, Li Z, Gu X, Xiang J. Colorectal cancer exosomes induce lymphatic network remodeling in lymph nodes. *Int J Cancer*. 2019;145(6):1648–59.
138. Pucci F, Garris C, Lai CP, Newton A, Pfirschke C, Engblom C, et al. SCS macrophages suppress melanoma by restricting tumor-derived vesicle-B cell interactions. *Science*. 2016;352(6282):242–6.
139. Chow A, Zhou W, Liu L, Fong MY, Champer J, Van Haute D, et al. Macrophage immunomodulation by breast cancer-derived exosomes requires Toll-like receptor 2-mediated activation of NF-kappaB. *Sci Rep*. 2014;4:5750.
140. Carrasco-Ramirez P, Greening DW, Andres G, Gopal SK, Martin-Villar E, Renart J, et al. Podoplanin is a component of extracellular vesicles that reprograms cell-derived exosomal proteins and modulates lymphatic vessel formation. *Oncotarget*. 2016;7(13):16070–89.
141. Biagioni A, Laurenzana A, Menicacci B, Peppicelli S, Andreucci E, Bianchini F, et al. uPAR-expressing melanoma exosomes promote angiogenesis by VE-Cadherin, EGFR and uPAR overexpression and rise of ERK1,2 signaling in endothelial cells. *Cell Mol Life Sci*. 2021;78(6):3057–72.
142. Zhou CF, Ma J, Huang L, Yi HY, Zhang YM, Wu XG, et al. Cervical squamous cell carcinoma-secreted exosomal miR-221-3p promotes lymphangiogenesis and lymphatic metastasis by targeting VASH1. *Oncogene*. 2019;38(8):1256–68.
143. Plebanek MP, Angeloni NL, Vinokour E, Li J, Henkin A, Martinez-Marin D, et al. Pre-metastatic cancer exosomes induce immune surveillance by patrolling monocytes at the metastatic niche. *Nat Commun*. 2017;8(1):1319.
144. Gajos-Michniewicz A, Duechler M, Czyz M. MiRNA in melanoma-derived exosomes. *Cancer Lett*. 2014;347(1):29–37.
145. Lee CK, Jeong SH, Jang C, Bae H, Kim YH, Park I, et al. Tumor metastasis to lymph nodes requires YAP-dependent metabolic adaptation. *Science*. 2019;363(6427):644–9.
146. Wong BW, Wang X, Zecchin A, Thienpont B, Cornelissen I, Kalucka J, et al. The role of fatty acid beta-oxidation in lymphangiogenesis. *Nature*. 2017;542(7639):49–54.
147. Zhang C, Yue C, Herrmann A, Song J, Egelston C, Wang T, et al. STAT3 activation-induced fatty acid oxidation in CD8(+) T effector cells is critical for obesity-promoted breast tumor growth. *Cell Metab*. 2020;31(1):148–61.
148. Howie D, Cobbold SP, Adams E, Ten Bokum A, Necula AS, Zhang W, et al. Foxp3 drives oxidative phosphorylation and protection from lipotoxicity. *JCI Insight*. 2017;2(3): e89160.
149. Mullen AR, DeBerardinis RJ. Genetically-defined metabolic reprogramming in cancer. *Trends Endocrinol Metab*. 2012;23(11):552–9.
150. Malhotra D, Fletcher AL, Astarita J, Lukacs-Kornek V, Tayalia P, Gonzalez SF, et al. Transcriptional profiling of stroma from inflamed and resting lymph nodes defines immunological hallmarks. *Nat Immunol*. 2012;13(5):499–510.
151. Wei WF, Chen XJ, Liang LJ, Yu L, Wu XG, Zhou CF, et al. Periostin(+) cancer-associated fibroblasts promote lymph node metastasis by impairing the lymphatic endothelial barriers in cervical squamous cell carcinoma. *Mol Oncol*. 2021;15(1):210–27.
152. Li Y, Wang L, Kang A, Fan L, Zhao Q, Tan B, et al. Screening and identification of proteins related to gastric cancer metastasis with comparative proteomics. *Nan Fang Yi Ke Da Xue Xue Bao*. 2015;35(3):360–4.
153. Tang B, Peng ZH, Yu PW, Yu G, Qian F. Expression and significance of Cx43 and E-cadherin in gastric cancer and metastatic lymph nodes. *Med Oncol*. 2011;28(2):502–8.
154. Fujita S, Sumi M, Tatsukawa E, Nagano K, Katase N. Expressions of extracellular matrix-remodeling factors in lymph nodes from oral cancer patients. *Oral Dis*. 2020;26(7):1424–31.
155. Allan AL, George R, Vantyghem SA, Lee MW, Hodgson NC, Engel CJ, et al. Role of the integrin-binding protein osteopontin in lymphatic metastasis of breast cancer. *Am J Pathol*. 2006;169(1):233–46.
156. Silvers CR, Messing EM, Miyamoto H, Lee YF. Tenascin-C expression in the lymph node pre-metastatic niche in muscle-invasive bladder cancer. *Br J Cancer*. 2021;125(10):1399–407.
157. Schudel A, Francis DM, Thomas SN. Material design for lymph node drug delivery. *Nat Rev Mater*. 2019;4(6):415–28.
158. He R, Zang J, Zhao Y, Dong H, Li Y. Nanotechnology-based approaches to promote lymph node targeted delivery of cancer vaccines. *ACS Biomater Sci Eng*. 2022;8(2):406–23.
159. Alexiou C, Arnold W, Klein RJ, Parak FG, Hulin P, Bergemann C, et al. Locoregional cancer treatment with magnetic drug targeting. *Cancer Res*. 2000;60(23):6641–8.
160. Alexiou C, Schmidt A, Klein R, Hulin P, Bergeman C, Arnold W. Magnetic drug targeting: biodistribution and dependency on magnetic field strength. *J Magn Magn Mater*. 2002;252:363–6.
161. Sasso MS, Mitrousis N, Wang Y, Briquez PS, Hauert S, Ishihara J, et al. Lymphangiogenesis-inducing vaccines elicit potent and long-lasting T cell immunity against melanomas. *Sci Adv*. 2021;7(13): eabe4362.
162. Francis DM, Manspecker MP, Schudel A, Sestito LF, O'Melia MJ, Kissick HT, et al. Blockade of immune checkpoints in lymph nodes through locoregional delivery augments cancer immunotherapy. *Sci Transl Med*. 2020;12: eaay3575.

## Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.